

For *In Vitro* Diagnostic Use

ANNUAL REVIEW

Reviewed by:	Date	Reviewed by:	Date

PRINCIPLE

INTENDED USE

VPA reagent, when used in conjunction with SYNCHRON LX[®] System(s), UniCel[®] DxH 600/800 System(s) and SYNCHRON[®] Systems Drug Calibrator 1 set, is intended for quantitative determination of valproic acid concentration in human serum or plasma.

CLINICAL SIGNIFICANCE

Valproic Acid is an anticonvulsant drug. It is indicated for the treatment of absence (petite mal), generalized tonic-clonic and myoclonic seizures. Valproic Acid therapy is monitored for suspected inadequate dose or toxicity.

METHODOLOGY

VPA reagent is used to measure the valproic acid concentration by a particle enhanced turbidimetric inhibition immunoassay method.¹ Particle-bound drug (PBD) binds to valproic acid specific antibody (Ab) resulting in the formation of insoluble aggregates causing turbidity. Non-particle-bound valproic acid in the patient sample competes with the PBD for the antibody binding sites, inhibiting the formation of insoluble aggregates. The rate and amount of particle aggregation is inversely proportional upon the concentration of valproic acid in the sample.

The SYNCHRON[®] System(s) automatically proportions the appropriate sample and reagent volumes into a cuvette. The ratio used is one part sample to 104 parts reagent. The system monitors aggregate formation by measuring the change in absorbance at 560 nanometers. This change in absorbance is inversely proportional to the concentration of valproic acid in the sample and is used by the SYNCHRON[®] System(s) to calculate and express the valproic acid concentration based upon a multi-point calibration curve.

CHEMICAL REACTION SCHEME



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SPECIMEN

TYPE OF SPECIMEN

Biological fluid samples should be collected in the same manner routinely used for any laboratory test.² Freshly drawn serum or plasma are the specimens of choice. Acceptable anticoagulants are listed in PROCEDURAL NOTES section of this chemistry information sheet. Whole blood or urine are not recommended for use as a sample.

SPECIMEN STORAGE AND STABILITY

1. Tubes of blood are to be kept closed at all times and in a vertical position. It is recommended that the serum or plasma be physically separated from contact with cells within two hours from the time of collection.³
2. Separated serum or plasma should not remain at room temperature longer than 8 hours. If assays are not completed within 8 hours, serum or plasma should be stored at +2°C to +8°C. If assays are not completed within 48 hours, or the separated sample is to be stored beyond 48 hours, samples should be frozen at -15°C to -20°C. Frozen samples should be thawed only once. Analyte deterioration may occur in samples that are repeatedly frozen and thawed.³

Additional specimen storage and stability conditions as designated by this laboratory:

SAMPLE VOLUME

The optimum volume, when using a 0.5 mL sample cup, is 0.3 mL of sample. For optimum primary sample tube volumes and minimum volumes, refer to the Primary Tube Sample Template for your system.

CRITERIA FOR UNACCEPTABLE SPECIMENS

Refer to the PROCEDURAL NOTES section of this chemistry information sheet for information on unacceptable specimens.

Criteria for sample rejection as designated by this laboratory:

PATIENT PREPARATION

Special instructions for patient preparation as designated by this laboratory:

SPECIMEN HANDLING

Special instructions for specimen handling as designated by this laboratory:

REAGENTS

CONTENTS

Each kit contains the following items:

Two Valproic Acid Reagent Cartridges (2 x 100 tests)

VOLUMES PER TEST

Sample Volume	3 µL
Total Reagent Volume	312 µL
Cartridge Volumes	
A	287 µL
B	--
C	25 µL

REACTIVE INGREDIENTS

REAGENT CONSTITUENTS

Valproic Acid Particle Reagent	4.1 mL
Monoclonal anti-Valproic Acid Antibodies (mouse)	43.6 mL

Also non-reactive chemicals necessary for optimal system performance.

CAUTION

Sodium azide preservative may form explosive compounds in metal drain lines. See National Institute for Occupational Safety and Health Bulletin: Explosive Azide Hazards (8/16/76).

MATERIALS NEEDED BUT NOT SUPPLIED WITH REAGENT KIT

SYNCHRON® Systems Drug Calibrator 1 set
At least two levels of control material
Saline

REAGENT PREPARATION

NOTICE

Failure to mix the reagent will result in erroneous values.

1. Gently invert the cartridge three times prior to loading onto the SYNCHRON System.
2. Check for bubbles or foam in compartments; break any bubbles.

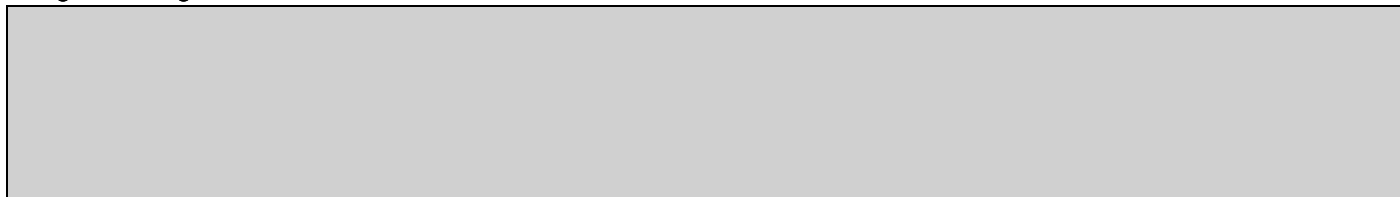
ACCEPTABLE REAGENT PERFORMANCE

The acceptability of a reagent is determined by successful calibration and by ensuring that quality control results are within your facility's acceptance criteria.

REAGENT STORAGE AND STABILITY

VPA reagent, when stored unopened at +2°C to +8°C, will remain stable until the expiration date printed on the cartridge label. Once opened, the reagent is stable for 42 days at +2°C to +8°C unless the expiration date is exceeded. Do not expose reagent to temperatures above +35°C or to direct sunlight.

Reagent storage location:



CALIBRATION

CALIBRATOR REQUIRED

SYNCHRON® Systems Drug Calibrator 1 set

CALIBRATOR PREPARATION

No preparation is required.

CALIBRATOR STORAGE AND STABILITY

SYNCHRON® Systems Drug Calibrator 1 set is stable until the expiration date printed on the calibrator bottle if stored capped in the original container at +2°C to +8°C.

CAUTION

Because this product is of human origin, it should be handled as though capable of transmitting infectious diseases. Each serum or plasma donor unit used in the preparation of this material was tested by United States Food and Drug Administration (FDA) approved methods and found to be negative for antibodies to HIV and HCV and nonreactive for HbsAg. Because no test method can offer complete assurance that HIV, hepatitis B virus, and hepatitis C virus or other infectious agents are absent, this material should be handled as though capable of transmitting infectious diseases. This product may also contain other human source material for which there is no approved test. The FDA recommends such samples to be handled as specified in Centers for Disease Control's Biosafety Level 2 guidelines.⁴

Calibrator storage location:

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CALIBRATION INFORMATION

1. The system must have a valid calibration curve in memory before control or patient samples can be run.
2. Under typical operating conditions the VPA reagent cartridge must be calibrated every 14 days and also with certain parts replacements or maintenance procedures, as defined in the SYNCHRON LX *Maintenance Manual and Instrument Log*, or the UniCel DxH 600/800 System *Instructions For Use* (IFU) manual. This assay has within-lot calibration available. Refer to the SYNCHRON LX *Operations Manual*, or the UniCel DxH 600/800 System *Instructions For Use* (IFU) manual for information on this feature.
3. For detailed calibration instructions, refer to the SYNCHRON LX *Operations Manual*, or the UniCel DxH 600/800 System *Instructions For Use* (IFU) manual.
4. The system will automatically perform checks on the calibration and produce data at the end of calibration. In the event of a failed calibration, the data will be printed with error codes and the system will alert the operator of the failure. For information on error codes, refer to the SYNCHRON LX *Diagnostics and Troubleshooting Manual*, or the UniCel DxH 600/800 System *Instructions For Use* (IFU) manual.

TRACEABILITY

For Traceability information refer to the Calibrator instructions for use.

QUALITY CONTROL

At least two levels of control material should be analyzed daily. In addition, these controls should be run with each new calibration, with each new reagent cartridge, and after specific maintenance or troubleshooting procedures as detailed in the appropriate system manual. More frequent use of controls or the use of additional controls is left to the discretion of the user based on good laboratory practices or laboratory accreditation requirements and applicable laws.

The following controls should be prepared and used in accordance with the package inserts. Discrepant quality control results should be evaluated by your facility.

Table 1.0 Quality Control Material

CONTROL NAME	SAMPLE TYPE	STORAGE

TESTING PROCEDURE(S)

1. If necessary, load the reagent onto the system.

2. After reagent load is completed, calibration may be required.
3. Program samples and controls for analysis.
4. After loading samples and controls onto the system, follow the protocols for system operations.

For detailed testing procedures, refer to the SYNCHRON LX *Operations Manual*, or the UniCel DxC 600/800 System *Instructions For Use* (IFU) manual.

CALCULATIONS

The SYNCHRON® System(s) performs all calculations internally to produce the final reported result. The system will calculate the final result for sample dilutions made by the operator when the dilution factor is entered into the system during sample programming.

REPORTING RESULTS

Equivalency between the SYNCHRON LX and UniCel DxC 600/800 Systems has been established. Chemistry results between these systems are in agreement and data from representative systems may be shown.

REFERENCE INTERVALS

Therapeutic VPA concentrations vary significantly, depending upon the individual. The lower limit for one patient may be ineffective in another, while the upper limit may prove toxic in a third. The physician should determine the appropriate reference interval for each patient. The reference intervals listed below were taken from literature.⁵

Table 2.0 Reference intervals

INTERVALS	SAMPLE TYPE	THERAPEUTIC INTERVAL		TOXIC INTERVAL	
		CONVENTIONAL UNITS (µg/mL)	S.I. UNITS (µmol/L)	CONVENTIONAL UNITS (µg/mL)	S.I. UNITS (µmol/L)
Literature	Serum/Plasma	50 – 100	347 – 693	> 100	> 693

INTERVALS	SAMPLE TYPE	THERAPEUTIC INTERVAL		TOXIC INTERVAL	
		CONVENTIONAL UNITS (µg/mL)	S.I. UNITS (µmol/L)	CONVENTIONAL UNITS (µg/mL)	S.I. UNITS (µmol/L)
Laboratory	Serum/Plasma				

Refer to References (6, 7, 8) for guidelines on establishing laboratory-specific reference intervals.

Additional reporting information as designated by this laboratory:

PROCEDURAL NOTES

ANTICOAGULANT TEST RESULTS

If plasma is the sample of choice, the following anticoagulants were found to be compatible with this method based on a study of 20 healthy volunteers:

Table 3.0 Acceptable Anticoagulants^a

ANTICOAGULANT	LEVEL TESTED FOR IN VITRO INTERFERENCE	AVERAGE PLASMA-SERUM BIAS (µg/mL)
Lithium Heparin	14 Units/mL	NSI ^b
Sodium Heparin	14 Units/mL	NSI

a Data shown was collected using SYNCHRON CX Systems. Equivalency between SYNCHRON LX Systems has been established by Deming regression analysis to SYNCHRON CX Systems.

b NSI = No Significant Interference (within ± 6.0 µg/mL or 10%).

LIMITATIONS

None identified.

INTERFERENCES

- The following substances were tested for interference with this methodology:

Table 4.0 Interferences^a

SUBSTANCE	SOURCE	LEVEL TESTED	OBSERVED EFFECT
Hemoglobin	RBC hemolysate	500 mg/dL	NSI ^b
Bilirubin	Bovine	30 mg/dL	NSI
Rheumatoid Factor	Human	300 IU/mL	NSI
Lipemia	Human	2+	NSI
Paraprotein (IgM)	Human	500 mg/dL	NSI

a Data shown was collected using SYNCHRON CX Systems. Equivalency between SYNCHRON LX Systems has been established by Deming regression analysis to SYNCHRON CX Systems.

b NSI = No Significant Interference (within ± 6.0 µg/mL or 10%).

- In very rare cases, patient samples may contain a particle agglutinating substance which may produce low results with this assay.
- Refer to References (9,10,11) for other interferences caused by drugs, disease and preanalytical variables.
- For assays employing mouse antibodies, the possibility exists for interference by human anti-mouse antibodies (HAMA) in the sample. Human anti-mouse antibodies may be present in samples from patients who have received immunotherapy or diagnostic procedures utilizing monoclonal antibodies or in individuals who have been regularly exposed to animals.^{12,13} Additionally, other heterophile antibodies, such as human anti-goat antibodies may be present in patient samples. Interpretation of results should be done in the context of the overall clinical presentation of the patient, including symptoms, clinical history, data from additional tests and other appropriate information.

SPECIFICITY

The following list of substances were added at the concentration listed to separate aliquots of a serum pool containing 50.0 µg/mL valproic acid. In most cases the value shown approximates maximum physiological concentrations.¹⁴ The recovered values were subtracted from the serum pool value. If the results were within $\pm 2X$ of the within-run precision specifications there was no significant interference. If the recovered results were more than $\pm 2X$ of the within-run precision specifications the difference is listed under observed effect.

Table 5.0 Specificity^a

SUBSTANCE	CONCENTRATION (µg/mL)	OBSERVED RECOVERY (µg/mL)	OBSERVED EFFECT
Carbamazepine	1000	48.31	NSI ^b
Clonazepam	100	54.01	NSI
Diazepam	100	51.30	NSI
Ethosuximide	1000	51.81	NSI
2-N-Propyl-2-pentenoic acid	100	53.91	NSI
2-N-Propyl-3-pentenoic acid	50	55.03	NSI
2-N-Propyl-4-pentenoic acid	10	53.83	NSI
3-Hydroxy-2-N-propylpentanoic acid	100	53.69	NSI
3-Hydroxy-2-N-propylpentanoic acid	10	51.52	NSI
4-Hydroxy-2-N-propylpentanoic acid	60	49.65	NSI
5-Hydroxy-2-N-propylpentanoic acid	25	52.22	NSI
3-Oxo-2-N-propylpentanoic acid	50	52.19	NSI
4-Oxo-2-N-propylpentanoic acid	15	54.04	NSI
2-(1'-Propenyl)-2-pentenoic acid	50	52.06	NSI
3-Oxo-2-(2'-Propenyl)-2-pentanoic acid	100	48.35	NSI
2-(2'-Propenyl)-4-pentenoic acid	25	52.19	NSI
Phenobarbital	750	50.52	NSI
PEMA = 2-Ethyl-2-phenylmalonamide	100	49.90	NSI
Phenytoin	1000	53.85	NSI
Primidone	1000	52.11	NSI
2-N-Propylglutaric acid	50	51.75	NSI
2-N-Propylmalonic acid	500	50.80	NSI
2-N-Propylsuccinic acid	100	54.64	NSI
Salicylic acid	3000	49.60	NSI
3-Ketovalproic acid	150	50.60	NSI

a Data shown was collected using SYNCHRON CX Systems. Equivalency between SYNCHRON LX Systems has been established by Deming regression analysis to SYNCHRON CX Systems.

b NSI = No Significant Interference (within ± 6.0 µg/mL or 10%).

PERFORMANCE CHARACTERISTICS

ANALYTIC RANGE

The SYNCHRON[®] System(s) method for the determination of this analyte provides the following analytical ranges:

Table 6.0 Analytical Range

SAMPLE TYPE	CONVENTIONAL UNITS	S.I. UNITS
Serum or Plasma	10.0 – 150.0 µg/mL	69 – 1040 µmol/L

Samples with concentrations outside of the analytical range will be reported as "<10.0 µg/mL" ("<69 µmol/L") or ">150.0 µg/mL" (">1040 µmol/L").

Samples reported out as greater than the analytical range may be confirmed by diluting with the valproic acid-free serum or plasma, saline, or the zero-level calibrator, and reanalyzing. The appropriate dilution factor should be applied to the reported result.

Samples reported out as less than the analytical range should be confirmed by diluting one part sample of known value with one part of the original patient sample. The assayed result of this dilution, when multiplied by 2, should approximate the original value of the known sample to confirm the low patient result. The confirmed result should be reported out as "<10.0 µg/mL" ("<69.0 µmol/L"). If the assayed result of the first dilution, when multiplied by 2, does not approximate the original result of the known sample, do not report result; assay by another method.

Samples reported out as "SUPPRESSED" due to RXN ERROR should be reanalyzed.

REPORTABLE RANGE (AS DETERMINED ON SITE):

Table 7.0 Reportable Range

SAMPLE TYPE	CONVENTIONAL UNITS	S.I. UNITS

SENSITIVITY

Sensitivity is defined as the lowest measurable concentration which can be distinguished from zero with 95% confidence. Sensitivity for this analyte determination is 10.0 µg/mL (69 µmol/L).

EQUIVALENCY

Equivalency was assessed by Deming regression analysis of patient samples to accepted clinical methods.

Serum or Plasma (in the range of 8.9 to 147.6 µg/mL):

Y (SYNCHRON LX Systems)	= 0.952X + 5.85
N	= 119
MEAN (SYNCHRON LX Systems)	= 66.3
MEAN (Fluorescence Polarization Immunoassay) ^a	= 63.5
CORRELATION COEFFICIENT (r)	= 0.989

^a A product of Abbott Laboratories, Abbott Park, IL.

Refer to References (15) for guidelines on performing equivalency testing.

PRECISION

A properly operating SYNCHRON® System(s) should exhibit precision values less than or equal to the following:

Table 8.0 Precision Values

TYPE OF PRECISION	SAMPLE TYPE	1 SD		CHANGEOVER VALUE ^a		% CV
		µg/mL	µmol/L	µg/mL	µmol/L	
Within-run	Serum/Plasma	3.6	25.0	60	416	6.0
Total	Serum/Plasma	4.5	31.2	60	416	7.5

a When the mean of the test precision data is less than or equal to the changeover value, compare the test SD to the SD guideline given above to determine the acceptability of the precision testing. When the mean of the test precision data is greater than the changeover value, compare the test % CV to the guideline given above to determine acceptability. Changeover value = (SD guideline/CV guideline) x 100.

Comparative performance data for a SYNCHRON LX[®] System evaluated using the NCCLS Proposed Guideline EP5-T2 appears in the table below.¹⁶ Each laboratory should characterize their own instrument performance for comparison purposes.

Table 9.0 NCCLS EP5-T2 Precision Estimate Method

TYPE OF IMPRECISION	SAMPLE TYPE		No. Systems	No. Data Points ^a	Test Mean Value (µg/mL)	EP5-T2 Calculated Point Estimates	
						SD	%CV
Within-run	Serum	Control 1	1	80	47.0	0.9	1.9
	Serum	Control 2	1	80	90.6	1.8	2.0
	Serum	Control 3	1	80	128.4	2.9	2.2
Total	Serum	Control 1	1	80	47.0	2.8	4.9
	Serum	Control 2	1	80	90.6	2.3	3.0
	Serum	Control 3	1	80	128.4	3.1	2.4

a The point estimate is based on the pooled data from one system, run for twenty days, two runs per day, two observations per run on an instrument operated and maintained according to the manufacturer's instructions.

NOTICE

These degrees of precision and equivalency were obtained in typical testing procedures on a SYNCHRON LX[®] System and are not intended to represent the performance specifications for this reagent.

ADDITIONAL INFORMATION

For more detailed information on SYNCHRON LX Systems or UniCel DxC Systems, refer to the appropriate system manual.

SHIPPING DAMAGE

If damaged product is received, notify your Beckman Coulter Clinical Support Center.

REFERENCES

1. Newman, D. J., Henneberry, H., Price, C. P., "Particle Enhanced Light Scattering Immunoassay", *Ann. Clin. Biochem.*, 29:22 42 (1992).
2. Tietz, N. W., "Specimen Collection and Processing; Sources of Biological Variation", *Textbook of Clinical Chemistry*, 2nd Edition, W. B. Saunders, Philadelphia, PA (1994).
3. National Committee for Clinical Laboratory Standards, *Procedures for the Handling and Processing of Blood Specimens*, Approved Guideline, NCCLS publication H18-A, Villanova, PA (1990).
4. CDC-NIH manual, *Biosafety in Microbiological and Biomedical Laboratories*, U.S. Government Printing Office, Washington, D.C. (1984).
5. Burtis, C. A., Ashwood, E. R., *Tietz Textbook of Clinical Chemistry*, 2nd Edition, W. B. Saunders, Philadelphia, PA (1994).
6. National Committee for Clinical Laboratory Standards, *How to Define, Determine, and Utilize Reference Intervals in the Clinical Laboratory*, Approved Guideline, NCCLS publication C28-A, Villanova, PA (1995).
7. Tietz, N. W., ed., *Fundamentals of Clinical Chemistry*, 3rd Edition, W. B. Saunders, Philadelphia, PA (1987).
8. Henry, J. B., *Clinical Diagnosis and Management by Laboratory Methods*, 18th Edition, W. B. Saunders Company, Philadelphia, PA (1991).
9. Young, D. S., *Effects of Drugs on Clinical Laboratory Tests*, 4th Edition, AACC Press, Washington, D. C. (1995).
10. Friedman, R. B., Young, D. S., *Effects of Disease on Clinical Laboratory Tests*, 3rd Edition, AACC Press, Washington, D.C. (1997).
11. Young, D. S., *Effects of Preanalytical Variables on Clinical Laboratory Tests*, 2nd Edition, AACC Press, Washington, D. C. (1997).
12. Bjerner, J., et al., "Immunometric Assay Interference: Incidence and Prevention", *Clin. Chem.* 48:613 621 (2002).
13. Kricka, L. J., "Interferences in Immunoassays-Still a Threat", *Clin. Chem.*, 46:1037 1038 (2000).
14. Siemes, H., Nau, H., Schultze, K., Wittfoht, W., Drews, E., Penzien, J., Seidel, U., "Valproate (VPA) Metabolites in Various Clinical Conditions of Probable VPA-Associated Hepatotoxicity", *Epilepsia*, 34(2):332 346, Raven Press, Ltd., New York, NY (1993).
15. National Committee for Clinical Laboratory Standards, *Method Comparison and Bias Estimation Using Patient Samples*, Approved Guideline, NCCLS publication EP9-A, Villanova, PA (1995).
16. National Committee for Clinical Laboratory Standards, *Precision Performance of Clinical Chemistry Devices*, Tentative Guideline, 2nd Edition, NCCLS publication EP5-T2, Villanova, PA (1992).

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